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Crepuscular and nocturnal illumination and its effects on color perception by the nocturnal hawkmoth *Deilephila elpenor*

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Summary

Recent studies have shown that certain nocturnal insect and vertebrate species have true color vision under nocturnal illumination. Thus, their vision is potentially affected by changes in the spectral quality of twilight and nocturnal illumination, due to the presence or absence of the moon, artificial light pollution and other factors. We investigated this in the following manner. First we measured the spectral irradiance (from 300 to 700 nm) during the day, sunset, twilight, full moon, new moon, and in the presence of high levels of light pollution. The spectra were then converted to both human-based chromaticities and to relative quantum catches for the nocturnal hawkmoth *Deilephila elpenor*, which has color vision. The reflectance spectra of various flowers and leaves and the red hindwings of *D. elpenor* were also converted to chromaticities and relative quantum catches. Finally, the achromatic and chromatic contrasts (with and

without von Kries color constancy) of the flowers and hindwings against a leaf background were determined under the various lighting environments. The twilight and nocturnal illuminants were substantially different from each other, resulting in significantly different contrasts. The addition of von Kries color constancy significantly reduced the effect of changing illuminants on chromatic contrast, suggesting that, even in this light-limited environment, the ability of color vision to provide reliable signals under changing illuminants may offset the concurrent threefold decrease in sensitivity and spatial resolution. Given this, color vision may be more common in crepuscular and nocturnal species than previously considered.

Key words: hawkmoth, *Deilephila elpenor*, nocturnal vision, color vision, environmental optics.

Introduction

While multiple visual pigments in certain deep-sea species (Cronin and Frank, 1996; Douglas et al., 1998) and multiple rod types in amphibians (Makino-Tasaka and Suzuki, 1984) have been known for some time, unambiguous evidence for true color vision under scotopic conditions has only recently been acquired (Kelber et al., 2002; Roth and Kelber, 2004). These behavioral studies, which show that the nocturnal hawkmoth *Deilephila elpenor* and the nocturnal helmet gecko *Tarentola chazaliae* can discern color under starlight and dim moonlight, respectively, raise at least two issues.

First, what is the selective advantage of color vision in these species that outweighs its costs? Color vision's detrimental effect on spatial resolution and the additional structural and neurological complexity required for color processing makes it a more difficult proposition for all species. However, color

vision presents additional difficulties for nocturnal species. While the decrease in sensitivity associated with the increase in the number of visual channels has little effect on species operating during light-saturated diurnal conditions, this sensitivity loss can potentially affect the ability of nocturnal species to function in their light-limited environment. It is primarily for this reason that color vision has generally been expected to be rare or absent among nocturnal species (Jacobs, 1993).

Second, what color are objects when viewed under the night sky? Although not perceived by humans, the spectrum of the night sky is not neutral, and depends on multiple factors, including how far the sun is below the horizon, the presence or absence and phase of the moon and, recently, on the level of light pollution (e.g. Munz and McFarland, 1977; Endler, 1991; Leinert et al., 1998; McFarland et al., 1999; Cinzano et al., 2001; Hernández-Andrés et al., 2001; Lee and Hernández-

Andrés, 2003). It has long been known that the variation of daytime spectra, due to cloud cover, solar elevation, forest canopy and depth (for aquatic species), has a substantial effect on the appearance and visibility of objects and organisms, which can be at least partly ameliorated by color vision (Wyszecki and Stiles, 1982; Endler, 1991; McFarland et al., 1999; Johnsen and Sosik, 2003; Lovell et al., 2005). Less work, however, has been done on the appearance of objects during twilight (reviewed by McFarland et al., 1999; Rickel and Genin, 2005), and, to our knowledge, the appearance of objects under different nocturnal illuminants has received very little attention.

This study measures or models spectral irradiance (300–700 nm) during daylight, sunset, twilight, moonlit nights, moonless nights and nights in regions with high light pollution. These spectra, in addition to previously published data, are then used to calculate the relative quantum catches of the three photoreceptors of *D. elpenor* under different lighting conditions. In addition to the general illuminants, relative quantum catches of five stimuli (green leaves, three flowers and the red hindwing of *D. elpenor*) are also calculated. Three different types of contrasts of the latter four stimuli viewed against green leaves are then determined: (1) achromatic contrast, (2) chromatic contrast and (3) chromatic contrast assuming von Kries color constancy. Finally, quantum catches of hypothetical photoreceptors with varying wavelengths of peak absorption are compared to the catches of the long wavelength receptor in *D. elpenor* under the different illuminants.

Materials and methods

General approach

The goal of this study was to determine the range of spectra found during sunset, twilight and night. Therefore, rather than measure a large number of spectra under all possible celestial and atmospheric conditions, we measured spectra under various extreme conditions. Both human-based chromaticities and the relative quantum catches described below have the property that the value of the mixture of two illuminants falls between the values of the two illuminants alone (Wyszecki and Stiles, 1982). Thus, by measuring the spectra under conditions where one of the various contributors to the illumination dominates, we can define the boundaries of the region where most spectra are found. The following four conditions were thus of particular interest: (1) clear nautical twilight (solar elevation between -6° and -12°), (2) full moon at high elevation under clear skies, (3) moonless and clear night and (4) urban overcast and moonless sky. The irradiances under these conditions correspond to nearly complete dominance by the following four factors respectively: (1) scattered sunlight modified by ozone absorption, (2) moonlight, (3) starlight and (4) anthropogenic illumination. These spectra were then compared with 2600 spectra of daylight, sunset and civil twilight (solar elevation between 0° and -6°) and 220 spectra of daylight under a forest

canopy using a model of color vision for the nocturnal hawkmoth, *Deilephila elpenor* L.

Measurement of twilight spectra

Fourteen sunset and twilight measurements of spectral irradiance under minimal cloud cover were taken on the beaches of two barrier islands located off the coast of North Carolina, USA (Atlantic Beach; $34^\circ 42' N$ $76^\circ 44' W$ and Cape Hatteras National Seashore; $35^\circ 44' N$ $75^\circ 32' W$, both at sea level) on 11 June, 12 June and 17 July, 2004. The locations were chosen to maximize the view of the sky and minimize the effects of anthropogenic light. Spectra were taken using a USB2000 spectrometer (Ocean Optics Inc., Dunedin, FL, USA) that had been modified for increased sensitivity by increasing the width of the entrance slit to 200 μm and focusing light onto the detector array with a collector lens (L2 collector lens, Ocean Optics). The spectrometer was fitted with a 1 mm diameter fiber optic cable that viewed a horizontal slab of a Lambertian reflector (Spectralon, Labsphere Inc., North Sutton, NH, USA). Because Lambertian materials reflect light evenly in all directions, their radiance is proportional to the irradiance striking them (Palmer, 1995). This method of obtaining the cosine response needed for measuring diffuse irradiance was chosen because it is more efficient than the typical diffusely transmitting disk (Doxaran et al., 2004).

Spectra were taken at solar elevations ranging from $+11^\circ$ to -11° (elevations determined using tables from the United States Naval Observatory). At lower solar elevations, the integration time of the spectrometer was increased to a maximum of 10 s, with 30 such integrations averaged per measurement. Spectra were taken from 300 to 700 nm and averaged over 5 nm intervals.

Measurement of full moonlight and synthesis of starlight spectra

Spectral irradiance under the full moon was measured using a spectrometer with a highly sensitive photomultiplier detector (OL-754-PMT, Optronics Laboratories Inc., Orlando, FL, USA). Spectra were taken on 10 December, 2003 at Harbor Branch Oceanographic Institution (Fort Pierce, FL, USA; $27^\circ 26' N$ $80^\circ 19' W$, sea level) during the full moon (elevation 69° , moon 98% full). An integrating sphere was used to ensure a cosine angular response. Data were taken at 5 nm intervals from 350 to 700 nm.

Preliminary attempts showed that even the OL-754 spectrometer was not sensitive enough to measure spectral irradiance on a moonless night. Therefore it was calculated in the following manner. The spectral radiances of small star-free portions of the moonless night sky were obtained from two observatories: Kitt Peak National Observatory (Tucson, AZ, USA; $31^\circ 58' N$ $111^\circ 36' W$, elevation 2083 m) and the William Herschel Telescope (La Palma, Canary Islands, Spain; $28^\circ 36' N$ $17^\circ 45' W$, elevation 2400 m) (Benn and Ellison, 1998; Massey and Foltz, 2000). Star and moon-free night spectra are composed primarily of airglow (emission spectra of the various molecular components of the upper atmosphere)

and zodiacal light (sunlight scattered from the dust in the plane of the solar system) (Leinert et al., 1997; Benn and Ellison, 1998). Because airglow is relatively constant over the entire hemisphere and zodiacal light is concentrated in a small region near the horizon, the former is the primary contributor to the diffuse irradiance of a star-free night sky (~80%) (Benn and Ellison, 1998). The stars contribute approximately 23–33% of the total irradiance, depending on the solar activity level (which affects the airglow intensity). The average spectrum of the stars of all spectral types (weighted by their relative abundances) was taken from Matilla (1980). This spectrum was combined with the star-free night sky spectra and integrated over the entire hemisphere of the sky to obtain estimates of the spectral irradiance on moonless nights. Two spectra were calculated from each observatory spectrum, one for the solar minimum (when stars contribute 33% of the total irradiance) and one for the solar maximum, (when stars contribute 23%). Spectra were calculated at 5 nm intervals from 300 to 700 nm.

To determine the effect of anthropogenic light on nocturnal irradiance, a spectrum was obtained from an urban location on a cloudy night (Jamaica Pond, Boston, MA, USA, 42°20'N 71°03'W, sea level) (M. Moore, unpublished data). Cloudy conditions were chosen because they maximize the effects of light pollution by reflecting urban lighting back to the ground. The measurement technique and resolution matched that described above for the North Carolina twilight spectra.

Daylight, civil twilight, and forest spectra

An estimate of the variability of daylight and civil twilight spectra (to compare with the variability during twilight and night) was obtained from 2395 daylight, 254 civil twilight and 220 forest measurements of spectral irradiance (Chiao et al., 2000; Hernández-Andrés et al., 2001; Lee and Hernández-Andrés, 2003). All the daylight and 205 of the civil twilight spectra were measured from the roof of the University of Granada's Science Faculty (Granada, Spain, 37°11'N 3°35'W, elevation 680 m) from February 1996 to February 1998 using a LI-1800 spectroradiometer (LI-COR Bioscience, Lincoln, NE, USA) fitted with a cosine-corrected receptor. Measurements were taken at all solar elevations greater than -4° and in all weather except for rain or snowfall. Data were collected at 5 nm intervals from 300 to 1100 nm. Another 49 civil twilight spectra were measured from three sites: Owings, MA, USA (38°41'N 76°35'W, elevation 15 m), Annapolis, MA, USA (38°59'N 76°29'W, elevation 18 m), and Marion Center, PA, USA (40°49'N 79°05'W, elevation 451 m). Measurements (from 380–780 nm) were taken from 1998 to 2001 using PR-650 spectroradiometer (Photo Research Inc., Chatsworth, CA, USA). Solar elevation ranged from 0° to -5.6° .

The 220 forest spectra were measured from sunrise to sunset during July and August 1999 in several temperate forests in Maryland, USA. Measurement locations included both full shade and under gaps in the canopy, and atmospheric conditions ranged from clear to overcast. Data were collected

at 3 nm intervals from 400 to 700 nm using an S2000 spectroradiometer (Ocean Optics) fitted with a cosine corrector.

UV and visible reflectance curves

The spectral reflectance of the white flower of the hawkmoth-pollinated evening primrose *Oenothera neomexicana* Munz (Raguso and Willis, 2002) and of the blue flower of the unspotted lungwort *Pulmonaria obscura* L. and the yellow flower of the birdsfoot trefoil *Lotus corniculatus* L. (Chittka et al., 1994) were used and are typical for white, yellow and blue flowers, respectively (although the flowers of certain species have higher reflectance at UV wavelengths). Reflections from a green leaf and the red area on the wings of the nocturnal hawkmoth *Deilephila elpenor* were measured using an S2000 Spectrometer (Ocean Optics) calibrated with a diffuse reflectance standard (WS1, Ocean Optics). All five spectra are shown in Fig. 1A.

Receptor sensitivities and photon catch calculation

The number of photons N that are absorbed by the

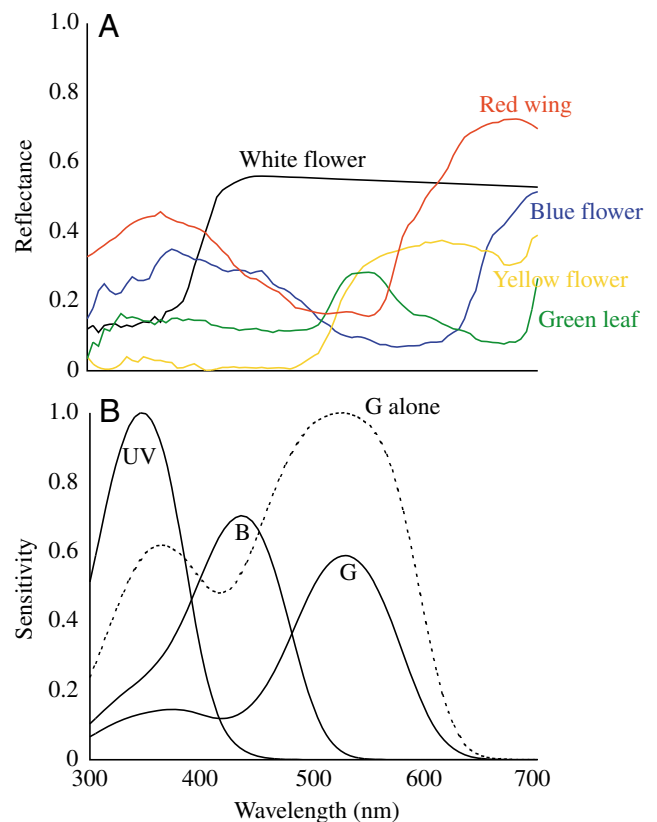


Fig. 1. (A) Spectral reflectance of stimuli (1=100%). (B) Spectral sensitivities of the photoreceptors of *Deilephila elpenor* assuming fused rhabdoms containing all three photoreceptor types. UV, B and G refer to the photoreceptors with peak absorption wavelengths of 350, 440, and 525 nm, respectively. Solid lines show normalized receptor sensitivities that were used to calculate relative quantum catches. The broken line shows the sensitivity of the green receptor that was used for the achromatic contrast calculations.

photoreceptors in one ommatidium of the nocturnal hawkmoth, *Deilephila elpenor*, per integration time of the photoreceptor, is given by:

$$N = 1.13 \left(\frac{\pi}{4} \right) n \Delta \rho^2 D^2 \Delta t \int \kappa \tau (1 - e^{-kR_i(\lambda)l}) L(\lambda) d\lambda \quad (1)$$

(Warrant and Nilsson, 1998; Kelber et al., 2002; Kelber et al., 2003a; Warrant, 2004). $L(\lambda)$ is the stimulus radiance in photons $\text{m}^{-2} \text{s}^{-1} \text{nm}^{-1} \text{sr}^{-1}$. $R_i(\lambda)$ ($i=1,2,3$) are the absorbance spectra of the three visual pigments of *D. elpenor*, calculated from their recorded sensitivity maxima (350 nm, 440 nm and 525 nm) (Schwemer and Paulsen, 1973; Höglund et al., 1973) using the Stavenga–Smits–Hoenders rhodopsin template (Stavenga et al., 1993) and equations 2a and 2b (Snyder et al., 1973). The other variables are given in Table 1.

For the calculation of the relative quantum catches, we assumed that the eyes of *D. elpenor* have fused rhabdoms with all three receptor types. This is a simplification because it is likely that there are two additional ommatidial types, one with blue and green receptors only, and one with UV and green receptors only (Kelber et al., 2002). However, because it is not known whether and how color processing involves inter-ommatidial connections, the ommatidial type containing all three receptors was the most general to model. Quantum catches were calculated assuming lateral screening (Snyder et al., 1973) (see Appendix for complete derivation). The receptor sensitivities were all normalized so that their integrals equalled 1. Thus, a stimulus that induces the same response in each photoreceptor type has its color locus in the centre of the color triangle (for details, see Kelber et al., 2003b).

Independent receptor adaptation was used as a model of chromatic adaptation (von Kries, 1904; Kelber et al., 2003b). This assumes that receptors adapt to the background intensity by keeping the response at approximately 50% of their maximal response (Laughlin and Hardie, 1978). The adapted receptor signal q is then:

$$q = N/N_b, \quad (2)$$

where N is quantum catch of a receptor viewing the stimulus and N_b is the quantum catch of the same receptor viewing the background. The radiance of the green leaves under the different illuminants was used as the background.

Table 1. *Optical and visual parameters for Deilephila elpenor*

Parameter	Description	Value
n	Effective facets in the superposition aperture	568
$\Delta\rho$	Photoreceptor acceptance angle	3.0°
D	Diameter of a facet lens	29 μm
κ	Quantum efficiency of transduction	0.5
τ	Fractional transmission of the eye media	0.8
Δt	Integration time of a photoreceptor	0.036 s
k	Absorption coefficient of the rhabdom	0.0067 μm^{-1}
l	Rhabdom length, doubled by tapetal reflection	414 μm

For calculating achromatic contrast, we assumed that green receptors extend over the entire length and width of the rhabdom and no lateral screening takes place (Fig. 1B, broken line). The achromatic contrast C was then calculated as:

$$C = \frac{N_x - N_{\text{green}}}{N_x + N_{\text{green}}}, \quad (3)$$

where N_x is the number of absorbed photons from the colored foreground and N_{green} is the number of absorbed photons from the green leaf background.

Because the spectra of nocturnal illumination are generally long-shifted (see Results), the 525 nm green pigment of *D. elpenor* may not be efficient at capturing this light. This possibility was examined by calculating the absolute photon catch of the long-wavelength pigment as a function of its peak wavelength. As was done for the achromatic contrast calculations, we assumed that green receptors extended over the entire length and width of the rhabdom and no lateral screening took place.

Results

Sunset, twilight and nocturnal spectra

Spectral irradiance changed substantially during sunset and twilight (Fig. 2A,B). As solar elevation decreased from 10° to 0°, the illumination gradually changed from being long-wavelength shifted to relatively spectrally neutral. After the disappearance of the sun's disk (thick line in Fig. 2A shows sunset), the spectra were dominated by a broad peak centered at ~450 nm, which became increasingly prominent as twilight progressed.

Nocturnal spectral irradiance was strongly affected by the presence or absence of the moon. Under a full moon at 70° elevation, the spectrum was nearly indistinguishable from a typical daylight spectrum. In the absence of the moon, the spectrum was shifted to longer wavelengths and displayed four narrow, but prominent peaks (at 560, 590, 630 and 685 nm). A moonless sky in a region with high amounts of light pollution was substantially long-wavelength shifted, with a broad peak centered at 590 nm.

Human-based chromaticity and relative quantum catches in D. elpenor

Mapping the twilight and nocturnal spectra into the perceptually uniform, human-based $u'v'$ chromaticity space showed that nautical twilight (solar elevation between -6° and -12°), moonless nights, and regions with high light pollution, had chromaticities well outside the envelope of those of the daylight, forest and early twilight illuminants (Fig. 3A). The same was also true for the relative quantum catches of *D. elpenor*, although the relative positions of starlight vs daylight vs twilight were different (Fig. 3B). The illumination of the full moon mapped to the long-wavelength border of the Granada daylight coordinates in both color spaces.

If humans had nocturnal color vision, these spectral shifts

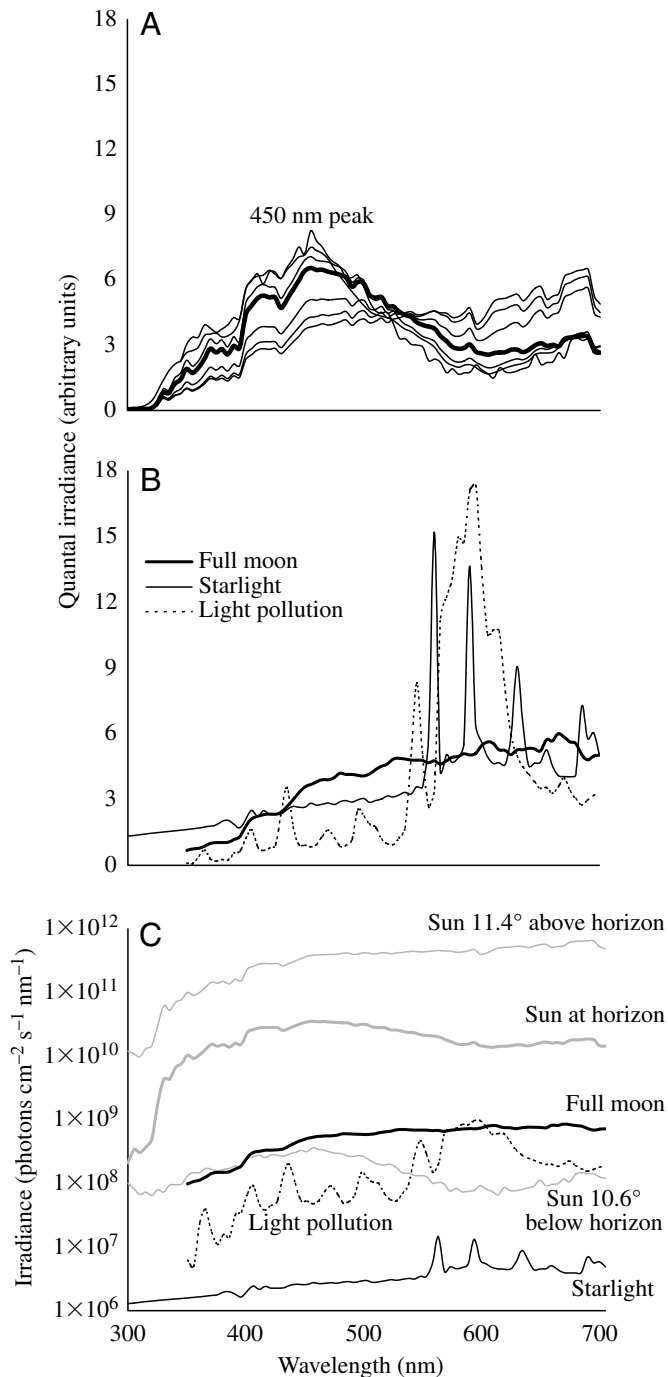


Fig. 2. (A) Normalized quantal irradiance during sunset and twilight. The thick line denotes a spectrum taken at sunset (solar elevation -0.6°). The three lines with long-wavelength irradiance greater than at sunset denote solar elevations of 11° , 5.9° and 1.0° (in order of decreasing long-wavelength values). The three lines with 450 nm peak values greater than at sunset denote solar elevations of -3.6° , -6.5° and -9.3° (in order of increasing 450 nm peak values). (B) Normalized quantal irradiance due to three common sources of nocturnal illumination. Spectra in A and B are normalized so that their irradiances integrated from 350 to 700 nm are all equal. (C) Unnormalized spectra. All spectra presented in this study, with the exception of those taken within forests, are freely available from the authors.

would be quite noticeable. When viewed under a light-polluted night, the evening primrose *Oenothera neomexicana* would appear far redder than under daylight. The same red shift, though smaller, would also be observed under starlight. When viewed under nautical twilight, the view would be strongly blue-shifted.

Relative quantum catches of flowers, leaves and wings

The relative quantum catches of the five examined stimuli (blue, white and yellow flowers; green leaves; red hindwings of *D. elpenor*) depended strongly on the source of illumination (Fig. 4A,B). In general, the variation was primarily in the relative quantum catch of the green photoreceptor (i.e. along a line connecting the green vertex to the UV–blue side). Decreasing solar elevation lowered the relative catch of the green receptor, with a slight increase in the relative catch of the UV receptor in nautical twilight. The type of nocturnal illumination affected the relative quantum catches to a similar degree, with all three illuminants (moonlight, starlight, light pollution) resulting in higher relative quantum catches in the green receptor. In general, the stimuli viewed under light-polluted skies had relative quantum catches substantially different from those under all natural illuminants, both crepuscular and nocturnal.

The variation of relative quantum catch was roughly similar among the five stimuli. The smallest and largest variations under twilight were found in the blue flower and green leaf stimuli respectively (Fig. 4B). The smallest and largest variations under the three nocturnal illuminants were found in the yellow flower and red wing stimuli respectively (Fig. 4A,B).

When von Kries color constancy was assumed, the variation of all five stimuli under the various illuminants was substantially less (Fig. 4C). The largest variation was found in the blue and yellow flower stimuli. The smallest variation was found in the red wing stimulus.

Achromatic and chromatic contrasts

The variation in achromatic contrast of the stimuli against the leaves under twilight, moonlight and starlight was strongly dependent on the stimulus (Fig. 5A,D). The achromatic contrast of the white flower stimulus was fairly independent of illuminant, with a coefficient of variation (i.e. standard deviation divided by the mean) of about 5%. In contrast, the achromatic contrasts of the yellow and blue flower stimuli had coefficients of variation higher than 100%. In addition, under full moon and starlight, their achromatic contrasts against the leaf were nearly zero. When the contrast of the two flowers under light polluted skies were also considered, the variation was even larger, with the contrasts switching polarities. The coefficient of variation of the red wing against the green leaves had an intermediate value of 27%.

In the case of chromatic contrasts (estimated as the distance between the relative quantum catches of the stimuli and the leaf background), the variation was in general lower and less dependent on stimulus, with coefficients of variation ranging

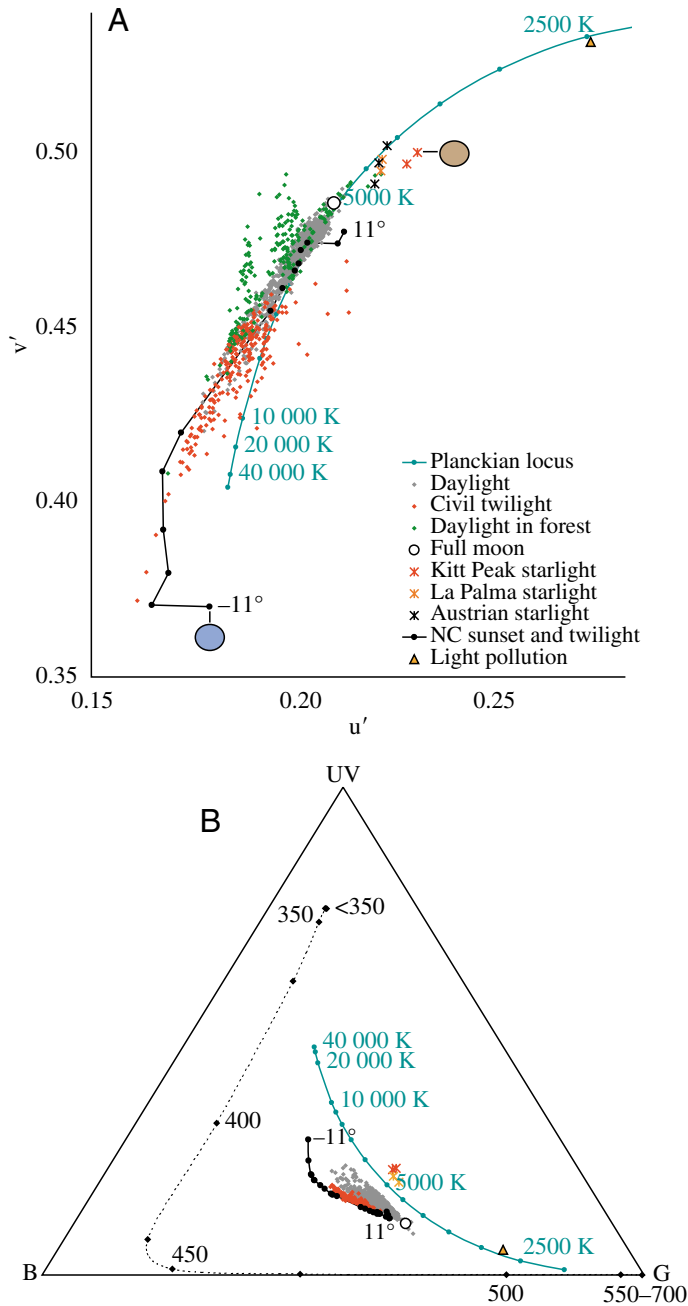


Fig. 3. (A) Human-based $u'v'$ chromaticities of daylight, sunset, twilight and nocturnal irradiances. The upper starlight symbols for the Kitt Peak and La Palma starlight data denote the chromaticities during a solar maximum; the lower symbols denote the chromaticities during a solar minimum. For comparison, the chromaticities of a 7° diameter patch of moonless sky (zenith angle 45°) under thin clouds, clear skies and overcast conditions are also shown (Höhn and Büchtermann, 1973). The black line denotes sunset and twilight data from North Carolina. Its symbols show data taken at solar elevation intervals of about 2° . The colored circles next to Kitt Peak starlight and -11° show the human-perceived colors at those two chromaticity extremes. The Planckian locus shows the chromaticities of blackbody radiators as a function of temperature. Data points for this locus are every 500 K up to 5000 K, and every 1000 K up to 10000 K, after which each point is labelled. (B) *Deilephila*-based relative quantum catches for the data shown in A. The three corners depict illuminants that are absorbed by one receptor only. The broken line shows the quantum catches of the spectral colors, with points every 25 nm and numbers every 50 nm. Because 49 of the civil twilight spectra and all 220 forest spectra were not taken at UV wavelengths, their relative quantum catches could not be calculated.

wavelengths. Under full moon and starlight though, the photon catch was strongly and positively correlated with λ_{\max} , with catches of hypothetical photoreceptors with 650 nm pigments being 2–3 times greater than those for the actual 525 nm long wavelength photopigment. This correlation was particularly strong for photoreceptors viewing the red wings of *D. elpenor* (Fig. 6C). The source of nocturnal illumination (starlight or moonlight) had little effect on this correlation for all three stimuli.

In fused rhabdoms containing all three pigments, the variation of the relative quantum catches under the three illuminants (given by the area of the triangle formed by the three quantum catch loci) also increased with wavelength. At peak wavelengths greater than 600 nm, the variation was 50% greater than it was at 525 nm.

Discussion

Spectral range of crepuscular and nocturnal illumination

Crepuscular and nocturnal periods provide challenging visual environments, where variations in intensity of up to six orders of magnitude co-occur with significant spectral variation. As the solar elevation decreases from $+20^\circ$ to -20° , the downwelling irradiance is first relatively spectrally neutral, then long-wavelength dominated, then short-wavelength dominated, and then either spectrally neutral or long wavelength dominated depending on the presence or absence of the moon (i.e. white to red to blue and then back to white or red, as perceived by humans). The same pattern in opposite order occurs at sunrise, and although not measured due to the limited sensitivities of the spectrometers, it is safe to assume that the same pattern also occurs at moonrise and moonset (given that moonlight is reflected sunlight). At temperate and tropical latitudes, the rate of solar and lunar elevation change near the horizon is approximately 1° every 4–6 min

from 14% to 36% without color constancy and from 1% to 24% assuming von Kries color constancy (Fig. 5B–D). Unlike for the achromatic case, chromatic contrasts were higher for the blue and yellow flower than for the white flower and red wing.

Photon catches as a function of the λ_{\max} of the long wavelength photoreceptor

Under nautical twilight, the photon catch of a receptor containing only one photopigment was relatively independent of the pigment's wavelength of peak absorption (λ_{\max}), regardless of whether the stimulus was white, green or red (Fig. 6A–C). However, there was a gradual decrease for hypothetical receptors with λ_{\max} at low visible and ultraviolet

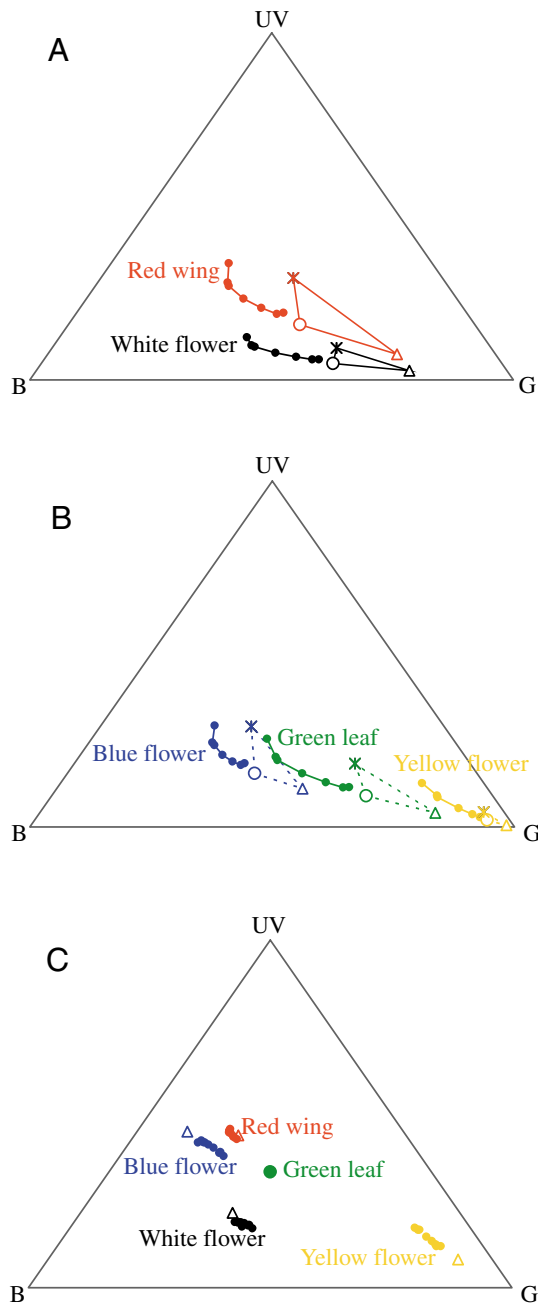


Fig. 4. (A,B) *Deilephila*-based relative quantum catches for the five different stimuli viewed under various sunset, twilight and nocturnal illuminants. Filled circles represent quantum catches of stimuli at sunset and twilight (solar elevation from 11° to -11°). Solar elevation decreases as data moves from right to left. Quantum catches under the nocturnal illuminants are to the right of those for sunset and twilight and consist of the following: open circles, quantum catches of stimuli under full moonlight; open triangles, quantum catches of stimuli under light polluted night sky; asterisks, quantum catches of stimuli under starlight only. (C) Quantum catches of the five stimuli assuming that *D. elpenor* has von Kries color constancy and is adapted to a background of green leaves under each illuminant (hence the central location of all the green stimuli). With the exception of light-polluted night skies (triangle), all the data have the same symbols for clarity.

(determined using US Naval Observatory tables). Thus, these intensity and spectral changes occur over a period of 2.5–4 h, with the central 20° range that exhibits the largest changes occurring in 80–120 min.

While cloud cover, solar elevation and the presence of a forest canopy also affect the spectral quality of daylight, the effect is smaller than what is observed during crepuscular periods and comparable to what is seen during the night. This is due partially to the fact that solar elevation has little effect on spectrum for elevations greater than 20° , and that clouds primarily scatter rather than absorb light, and thus have little effect on spectral quality. More important, however, is that the only two significant sources of daytime illumination are the sun and scattered sunlight, whose spectral characteristics and relative contributions both remain fairly constant at solar elevations greater than 20° . In contrast, crepuscular and nocturnal environments are lit by multiple sources with different spectra including a low-elevation sun or moon, high elevation moon, starlight, airglow emissions, and scattered sun or moonlight (Leinert et al., 1998). Because both the intensities and spatial extents of these sources vary by many orders of magnitude (Fig. 2C), spectral quality can change rapidly and significantly, particularly during the rising and setting of the sun or moon (Fig. 7). For example, near sunset the small, but intense and long-wavelength dominated solar disk balances the relatively dim short-wavelength dominated skylight until the sun nearly reaches the horizon, after which the general illumination changes rapidly from spectrally neutral to short-wavelength dominated.

Surprisingly, the intense blue of skylight during nautical twilight is not due to wavelength-dependent light scattering, but to absorption by ozone (Hulbert, 1953; Rozenberg, 1966). In addition to its strong absorption at ultraviolet wavelengths, ozone also has a broad absorption band in the visible, known as the Chappuis Band. While this absorption has only a minor effect on the spectrum of the daytime sky, it has a profound effect during late twilight. Without this absorption, which ranges from 450 to 700 nm and has double peaks at approximately 580 and 600 nm, skylight during nautical twilight would be a pale yellow (reviewed by Bohren, 2004). Because ozone concentration varies with season, geographic location and human activity (reviewed by Vingarzan, 2004), the spectra of skylight during nautical twilight are likely to be quite variable.

Changing crepuscular and nocturnal illumination and monochromatic visual systems

Although the exact achromatic contrasts depend on the spectral sensitivity of the viewer and the spectral reflectances of the targets and backgrounds, the examples given in this study show that they can vary significantly under the different crepuscular and nocturnal illuminants. With the exception of the white flower, the achromatic contrasts of the stimuli against the leaf background were quite variable. In certain cases, the contrast changed polarity. For example, the blue flower was brighter than the leaves during nautical twilight, but darker

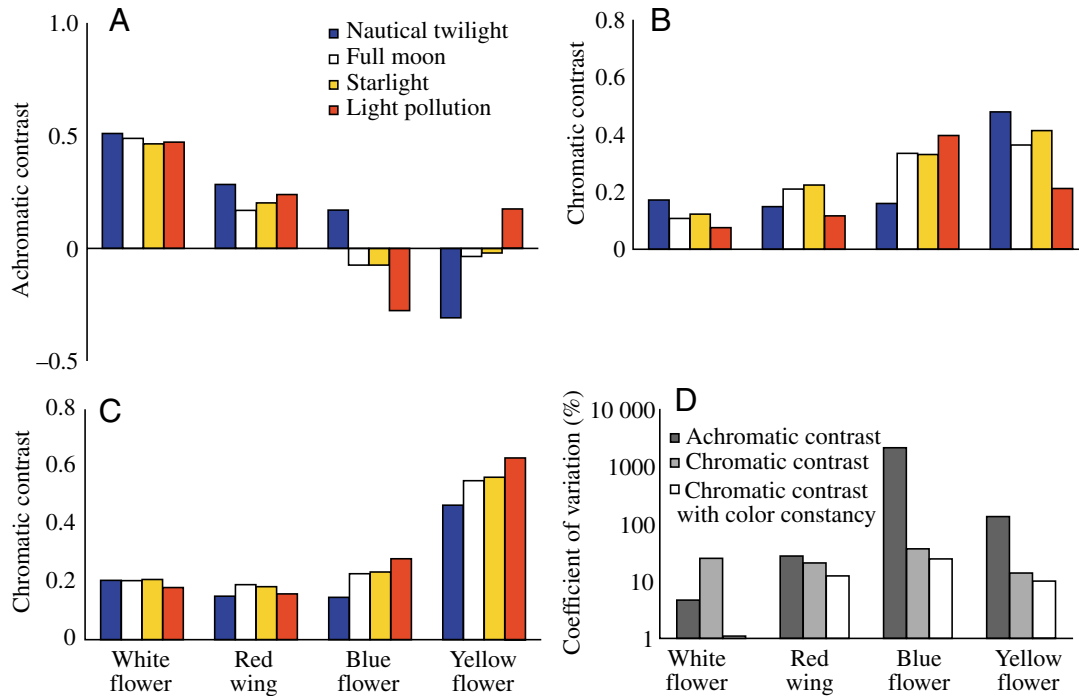


Fig. 5. (A) Achromatic contrast between four stimuli (the white evening primrose, the red hindwing of *D. elpenor*, the yellow flower, and the blue flower) and a green leaf background. Positive contrast indicates that the object is brighter than the background. (B) Chromatic contrast, defined as the distance between the relative quantum catches of the stimuli and the leaf background. (C) Chromatic contrast assuming that *D. elpenor* has von Kries color constancy and is adapted to a background of green leaves under each illuminant. (D) Coefficients of variation of the achromatic and chromatic contrasts of the four stimuli when viewed under nautical twilight, moonlight and starlight.

than the leaves during night. In addition, two of the stimuli (the blue and yellow flowers) had low contrasts under moonlight and starlight, likely rendering them undetectable *via* achromatic cues.

In contrast, the white flower, whose reflectance is high but relatively similar in spectrum to the leaves, had a high and stable contrast under all light conditions (Fig. 5A). *D. elpenor* and other nocturnal hawkmoths are thought to primarily visit white flowers with exceptionally high reflectance (reviewed by Raguso and Willis, 2002; Kelber et al., 2003a). In addition, crepuscular hawkmoths (e.g. *Manduca sexta*), and those that are active both during day and night (e.g. *Hyles* sp.), tend to visit blue and yellow flowers in bright light but white flowers in dim light (reviewed by Raguso and Willis, 2002).

The need for stability of achromatic contrast may also explain why the nocturnal flowers of many bat-pollinated species tend to be red or white. Flower-visiting bats are colorblind at night (Winter et al., 2003) and thus rely on achromatic contrast. Because the illumination during moonlit and starlit nights is long-wavelength shifted, red flowers are bright relative to green leaves, resulting in a high and more stable contrast. However, because the peak wavelength of the long-wavelength pigments of some of these bats is relatively low (~510 nm), they may not be able to exploit this contrast.

In general, however, achromatic contrast depends strongly on the illuminant, which varies significantly during crepuscular and nocturnal periods. This variation, which occurs whenever

spectrally different stimuli and backgrounds are viewed under highly variable illuminants, makes monochromatic vision unreliable during these periods.

Chromatic contrasts and color constancy

While chromatic contrasts varied less than achromatic contrasts (Fig. 5D), the addition of color constancy, which has recently been demonstrated for *D. elpenor* (Balkenius and Kelber, 2004), reduces the variation further. Chromatic contrasts without constancy are affected by the fact that the different lighting conditions changed the relative quantum catches from different colored stimuli in different ways. For example, relative quantum catches from the yellow flower *Lotus corniculatus* viewed under moonlight and nautical twilight changed less than did the relative quantum catches from the green leaf background (Fig. 4B). This is due to the fact that the relative contribution of the long-wavelength light that the yellow flower reflects changes less than the relative contribution of the middle wavelength light that the leaf reflects (Figs 1A, 2A,B). The result is not only a shift in the color of the scene, but also of the chromatic contrast between the flower and the leaf background. Color constancy, which can be explained as the result of receptor adaptation, reduces the variation for all four stimuli. In the case of the white flower, whose variation in chromatic contrast is greater than its variation in achromatic contrast, color constancy removes nearly all the variation.

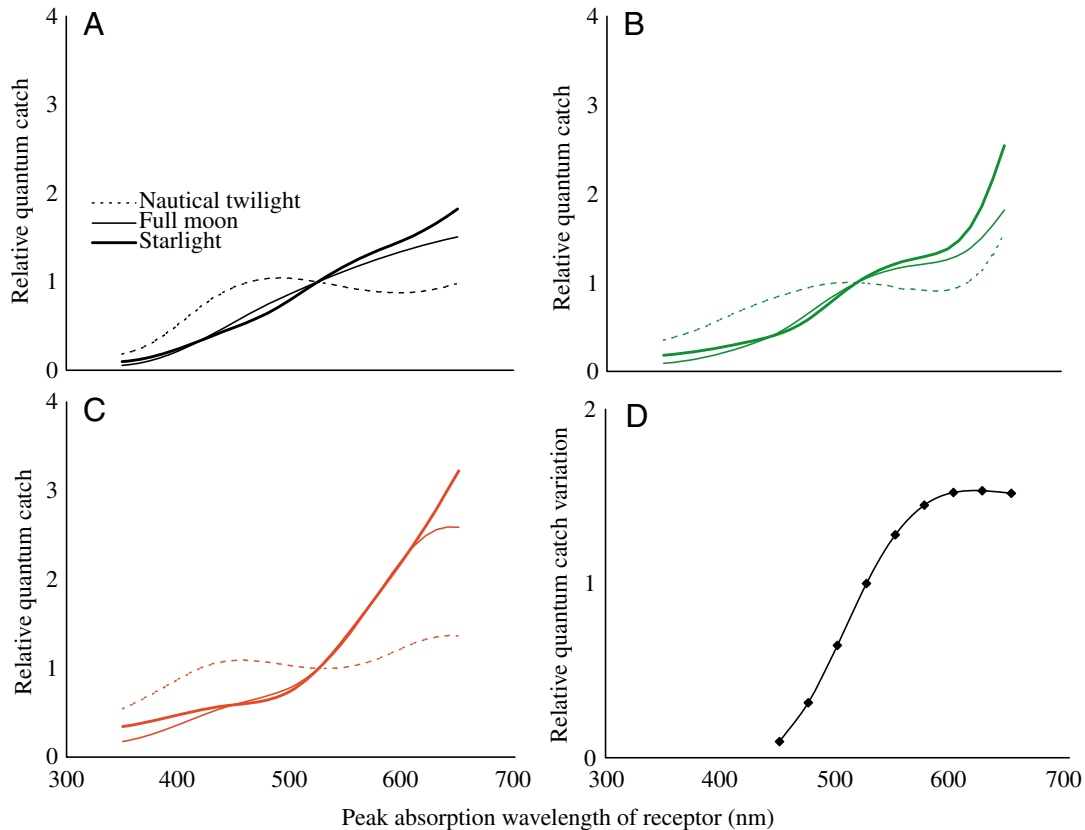


Fig. 6. (A–C) Numbers of photons absorbed by a hypothetical photoreceptor with a given λ_{\max} relative to the number absorbed by the green receptor possessed by *D. elpenor* ($\lambda_{\max}=525$ nm) under three illuminants (nautical twilight, full moon, starlight). (A) Viewing the white evening primrose. (B) Viewing green leaves. (C) Viewing the red hindwing of a conspecific. (D) The variation in relative quantum catches (among nautical twilight, moonlight and starlight), in a fused rhabdom containing all three visual pigments, as a function of the wavelength of the long wavelength receptor. The variation is estimated by the area of the triangle formed by the three points in the Maxwell triangle. As in A–C, the variation at 525 nm is set to 1.

The function of nocturnal color vision in *D. elpenor* is poorly understood. As mentioned above, nocturnal hawkmoths are thought to visit white flowers at night, which can reliably be detected without color vision. However, given that other hawkmoths visit blue and yellow flowers during the day, it is possible that flowers of these colors are also visited at night. Given their unreliable appearance to monochromatic visual systems, blue and yellow flowers may remain undetected by competitors of *D. elpenor*, allowing them to exploit an additional source of nectar.

The general long-wavelength shift of nocturnal illumination and the red coloration of *D. elpenor* render this species quite visible at night. Also, it has relatively stable achromatic and chromatic contrasts (Fig. 5). While many hawkmoths have some red coloration, particularly on their hindwings (which is thought to function as a startle display), the more extensive red coloration of *D. elpenor* is less common (Kitching and Cadiou, 2000). This raises

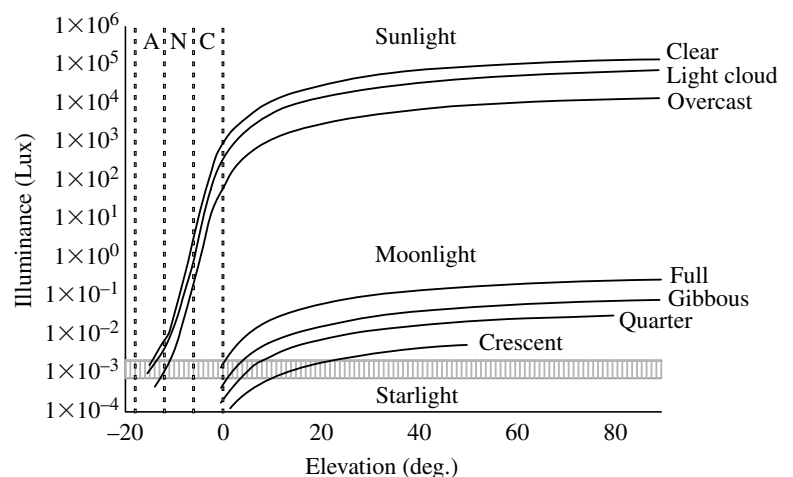


Fig. 7. The variation of sunlight and moonlight relative to starlight as a function of the elevation of the sun or moon, the sky conditions, and the phase of the moon. A, N and C refer to astronomical, nautical and civil twilight, respectively. (Modified from Bond and Henderson, 1963.)

the possibility that color vision may enhance recognition of conspecifics or be used in mating. While mating in moths is thought to be entirely mediated by olfaction, most tasks are eventually found to involve multiple sensory modalities. For example, nocturnal foraging in hawkmoths is known to involve both visual and olfactory cues (Raguso and Willis, 2002).

Absolute numbers of captured photons as a function of λ_{max}

While the relationship between visual pigment maxima and illuminant spectra under diurnal conditions is complex, research on deep-sea fish has shown that, at least in that particular light-limited environment, visual sensitivity peaks close to the wavelength of peak illumination (reviewed by Partridge and Cummings, 1999). This characteristic, which maximizes photon catch, does not appear to operate in *D. elpenor*. The peak wavelength is similar to those found in the long wavelength receptors of diurnal moths (Briscoe and Chittka, 2001), and differs substantially from that leading to maximal photon catch (Fig. 6A–C). This is intriguing, given the extreme light limitation present during color vision under starlight (Kelber et al., 2002), and the presence of longer wavelength pigments in the Lepidoptera (Briscoe and Chittka, 2001). In vertebrates, the higher noise levels in long-wavelength ciliary receptors (dark noise) (Barlow, 1957; Donner et al., 1990; Firsov and Govardovskii, 1990; Ala-Laurila et al., 2004) may account for this. However, dark noise appears to play a minor role in invertebrates due to different transduction mechanisms in rhabdomeric receptors (Laughlin, 1990; Warrant, 2004). Because relative quantum catches in the fused rhabdoms of *D. elpenor* vary more with changing illuminant as the peak wavelength of the long-wavelength receptor increases (Fig. 6C), the 525 nm peak may be a compromise between sensitivity and color stability. The peak wavelengths of the photoreceptors may also be constrained by their function during diurnal periods.

It is also possible that the sub-optimal λ_{max} of the long wavelength pigment is due to a phylogenetic or other constraint. Indeed, a survey of visual pigment maxima in insects by Briscoe and Chittka (2001) found little correlation with environment or behavior. However, at least two nocturnal species in the moth family Noctuidae have a fourth visual pigment (λ_{max} =560, 580 nm) (Langer et al., 1979; Ichikawa and Tateda, 1982), which phylogenetic analyses suggest are independently evolved within the Lepidoptera (Briscoe and Chittka, 2001). The function of these pigments at nocturnal light levels is doubtful given the limited optical sensitivity of noctuid eyes (A. Kelber, unpublished data), but their existence casts some doubt on a phylogenetic constraints argument.

Light pollution

Anthropogenic light sources ('light pollution') are an increasingly dominant factor in nocturnal illumination (e.g. Cinzano et al., 2001; Garstang, 2004). In addition to reaching intensities comparable to the light during nautical twilight or

under the full moon, spectral irradiance under light polluted skies is substantially different from that found under any natural illumination (Fig. 3). While light pollution spectra have many peaks (primarily due to mercury and sodium emission lamps), the primary spectral difference is a large increase in the relative contribution of long-wavelength light. This significantly changed both the achromatic and chromatic contrasts of the considered stimuli. The achromatic contrasts of the blue and yellow flowers in particular were significantly altered.

Light pollution can rival the intensity of the blue sky during nautical twilight and essentially has an opposite spectrum: the former being strongly long-wavelength shifted, the latter strongly short-wavelength shifted. Therefore the color of twilight illumination in urban and other light-polluted regions will vary rapidly over an unnaturally large range, potentially presenting significant difficulties for both monochromatic and color-visual species operating during this period.

Recent research on the ecological effects of light pollution (reviewed by Longcore and Rich, 2004) has generally focused on its intensity. To our knowledge, however, no studies have examined the effect of the color of light pollution. Given its unusual spectrum, it may have a significant effect on the foraging and mating of crepuscular and nocturnal species.

Conclusions

The spectral quality of crepuscular and nocturnal illumination varies over a larger range than does that of diurnal illumination, even when a wide range of atmospheric and forest conditions are considered. This variation makes monochromatic visual systems unreliable during these periods. We propose, for species that forage during twilight and night, that the increased signal reliability afforded by color constant color vision offsets the decreased sensitivity and provides an explanation for this unusual trait. However, the preference of *D. elpenor* for white flowers, which have stable achromatic contrasts, complicate the picture for this species. The mismatch of the long-wavelength pigment to the spectra of nocturnal illumination results in a less than optimal photon catch, but may lead to higher color stability. Light polluted night skies are strongly long-wavelength shifted and substantially alter the appearance of objects. Future research into nocturnal vision will need to consider the large natural and anthropogenic variability of this optical environment.

Appendix

Calculation of relative quantum catches assuming a fused rhabdom containing equal volumes and cross-sectional areas for each photopigment

$L(\lambda)$ is the stimulus strength (in quanta) at distal surface of the rhabdom; $R_i(\lambda)$ is the absorbance curve of i th pigment, where i =UV, B or G, normalized to a peak of 1; $\bar{R}(\lambda)$ is the un-normalized average of the three absorbance curves; and k and l are absorption coefficient and length of the rhabdom, respectively.

The number of photons of wavelength λ that penetrate a distance x into the rhabdom equals:

$$L(\lambda)e^{-k\bar{R}(\lambda)x}. \quad (\text{A1})$$

The fraction of these photons that are absorbed by a dx thick section of the portion of the rhabdom containing the i th photopigment equals:

$$\frac{1 - e^{-kR_i(\lambda)dx}}{3} \cong \frac{kR_i(\lambda)}{3} dx \quad (\text{A2})$$

(from Taylor expansion of e^x for small x).

Thus, the total number of photons absorbed by the i th photopigment at wavelength λ by the entire rhabdom equals:

$$Q_i(\lambda) = \frac{1}{3} L(\lambda)kR_i(\lambda) \int_0^l e^{-k\bar{R}(\lambda)x} dx = \left| -\frac{1}{3} L(\lambda) \frac{kR_i(\lambda)}{k\bar{R}(\lambda)} e^{-k\bar{R}(\lambda)x} \right|_0^l \quad (\text{A3})$$

Evaluating this integral at l and 0 gives:

$$Q_i(\lambda) = -\frac{1}{3} L(\lambda) \frac{R_i(\lambda)}{\bar{R}(\lambda)} e^{-k\bar{R}(\lambda)l} + \frac{1}{3} L(\lambda) \frac{R_i(\lambda)}{\bar{R}(\lambda)} = \frac{1}{3} L(\lambda) \frac{R_i(\lambda)}{\bar{R}(\lambda)} (1 - e^{-k\bar{R}(\lambda)l}). \quad (\text{A4})$$

Therefore, the total quantum catch by the i th photopigment is:

$$Q_i = \frac{1}{3} \sum_{\lambda=300 \text{ nm}}^{\lambda=700 \text{ nm}} L(\lambda) \frac{R_i(\lambda)}{\bar{R}(\lambda)} (1 - e^{-k\bar{R}(\lambda)l}) \Delta\lambda. \quad (\text{A5})$$

The color locus of a given stimulus $L(\lambda)$ is (X_1, X_2) , where:

$$X_1 = \frac{1}{\sqrt{2}} (q_G - q_B), \quad \text{and} \quad X_2 = \frac{\sqrt{2}}{\sqrt{3}} \left(q_{UV} - \frac{q_G + q_B}{2} \right), \quad (\text{A6})$$

where:

$$q_{UV} = \frac{Q_{UV}}{Q_{UV} + Q_B + Q_G}, \quad q_B = \frac{Q_B}{Q_{UV} + Q_B + Q_G}, \quad q_G = \frac{Q_G}{Q_{UV} + Q_B + Q_G}. \quad (\text{A7})$$

Before the calculation of the relative quantum catches, the Q_i values are normalized so that $Q_{UV} = Q_B = Q_G$ for any spectrally neutral (e.g. color-less) stimulus. This maps these stimuli to the center of the color triangle. This normalization is done by dividing each Q_i by

$$\sum_{\lambda=300 \text{ nm}}^{\lambda=700 \text{ nm}} L(\lambda) \frac{R_i(\lambda)}{\bar{R}(\lambda)} (1 - e^{-k\bar{R}(\lambda)l}). \quad (\text{A8})$$

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Commentary

Nocturnal colour vision – not as rare as we might think

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Summary

The dual retina of humans and most vertebrates consists of multiple types of cone for colour vision in bright light and one single type of rod, leaving these animals colour-blind at night. Instead of comparing the signals from different spectral types of photoreceptors, they use one highly sensitive receptor, thus improving the signal-to-noise ratio. However, nocturnal moths and geckos can discriminate colours at extremely dim light intensities when humans are colour-blind, by sacrificing spatial and temporal rather than spectral resolution. The advantages of colour vision are just as obvious at night as

they are during the day. Colour vision is much more reliable than achromatic contrast, not only under changing light intensities, but also under the colour changes occurring during dusk and dawn. It can be expected that nocturnal animals other than moths and geckos make use of the highly reliable colour signals in dim light.

Key words: vision, colour vision, sensitivity, colour constancy, night vision.

Preconditions for colour vision

Colour vision is commonly defined as the ability to discriminate between two visual stimuli that only differ in their spectral composition, independent of their relative intensities (e.g. Kelber et al., 2003b). A large number of animals are known to use colour to detect, discriminate and recognise objects such as food sources (for instance flowers or fruit), mating partners (as is the case in butterflies and birds), landmarks or their homes. For colour vision to be possible, an animal needs to possess and use at least two types of photoreceptors, with different spectral sensitivities, to look at an object (stage 1 in Fig. 1A). The signals from these receptors need to be compared by neurons in the visual system (stage 2 in Fig. 1A) to produce a chromatic or colour signal. Alternatively, receptor signals can be summed to produce an achromatic (intensity-related) signal. In human colour vision, brightness refers to the achromatic, and hue and saturation to the chromatic, properties of a colour. For colour discrimination, the signals arising from two different stimuli must then be compared (stage 3 in Fig. 1A) and result in a behavioural reaction in the animal (stage 4 in Fig. 1A). Most animals that possess multiple types of photoreceptors use them for colour discrimination (for references, see Kelber et al., 2003b).

Many invertebrates possess two (cockroaches, ants), three (giant clams, firefly squid, bees, wasps, moths, hunting spiders), four (water fleas), five (flies and some butterflies) or

even twelve (stomatopod crustaceans) types of receptor that can be used for colour vision (for references, see Briscoe and Chittka, 2001; Cronin and Hariyama, 2002; Kelber, 2006). However, not all of the receptors are used for colour vision. Flies, a well-studied example, use only four out of their five receptor types for colour vision; the fifth receptor has a broad spectral sensitivity and is used exclusively for achromatic vision, as is obvious from the complete separation of both visual pathways (Strausfeld and Lee, 1991; Osorio and Vorobyev, 2005; Kelber, 2006). The firefly squid has three spectral types of receptor but only two of them are localized in the same part of the retina (Michinomae and Masuda, 1994).

In vertebrates, the situation is somewhat more complicated. Most of them possess a dual retina consisting of rods used for vision in dim light and cones used for vision in bright light. Only cones make a major contribution to colour vision. At scotopic light intensities (below 0.005 cd m^{-2} , see Fig. 1B) only rods contribute to (colour-blind) vision, and at photopic light levels (above 5 cd m^{-2}), most vertebrates including humans use only cones to see colour. At mesopic light levels both cones and rods contribute and colours look less saturated (Ambler and Proctor, 1976).

Some vertebrates have pure rod retinæ (e.g. Douglas et al., 1998), but most have one type of rod and one (marine and some nocturnal mammals), two (the majority of mammals), three

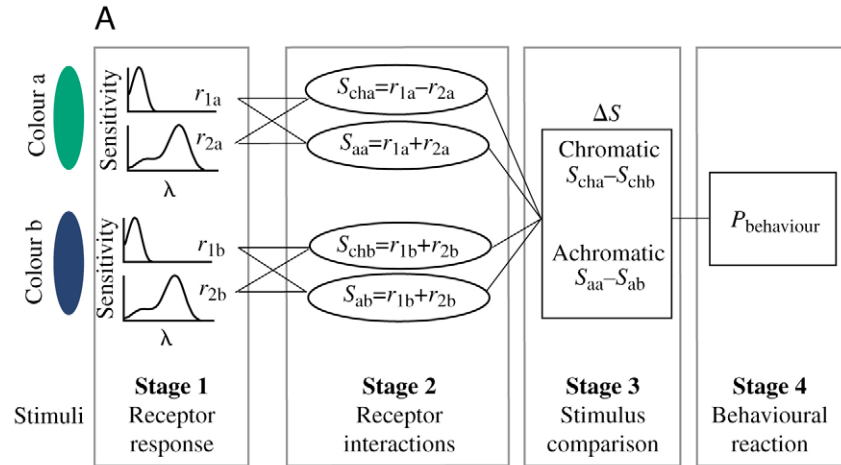
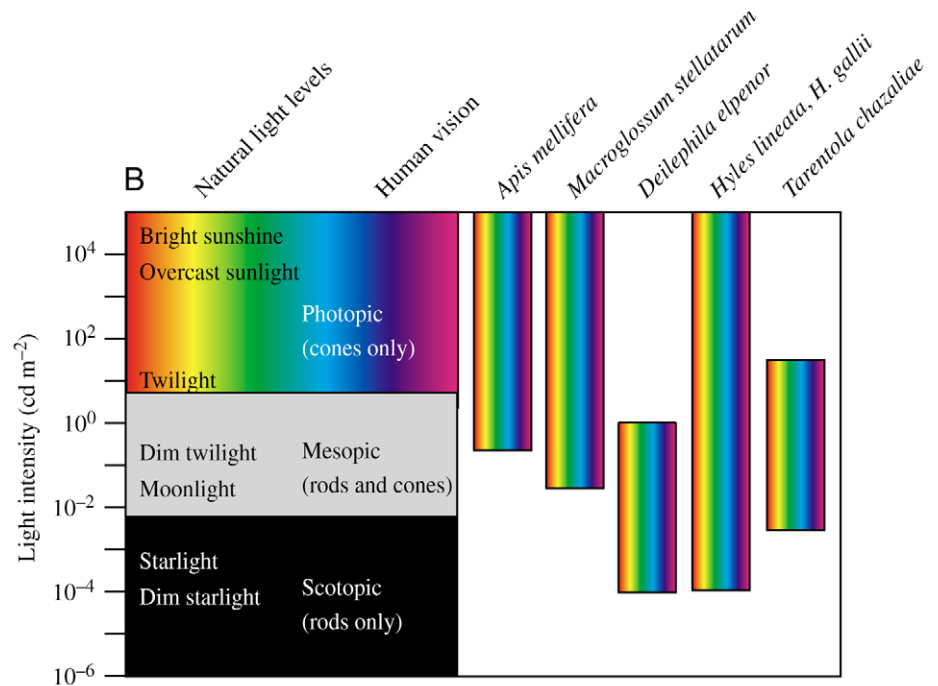


Fig. 1. (A) Simple 4-stage model of colour discrimination with two spectral types of receptors. At receptor stage 1, the signals r_{1a} , r_{1b} and r_{2a} , r_{2b} arise when the animal looks at the two colours a and b. At the subsequent neural stage 2, two neural interactions are possible: summation of the receptor signals, resulting in the achromatic signals S_{aa} and S_{ab} , and subtraction (or comparison), resulting in the chromatic signals S_{cha} and S_{chb} . At stage 3, the signal arising from the two colours a and b are compared, and finally, a behaviour will occur with probability P . (Adapted from Kelber et al., 2003b.) (B) Natural light levels and limits of colour vision in different animals. Humans lose their colour vision ability in dim moonlight and so do diurnal honeybees *Apis mellifera* (Menzel, 1981). Nocturnal hawkmoths (*Deilephila elpenor*, *Hyles lineata* and *H. gallii*) can still see colour at dim starlight levels. Nocturnal geckos (*Tarentola chazaliae*) were tested at dim moonlight levels.



(primates, and some fish, amphibians and reptiles) or four (some fish, most reptiles and birds) types of cone (for a review see Kelber et al., 2003b). Lizards have pure cone retinæ (Underwood, 1970).

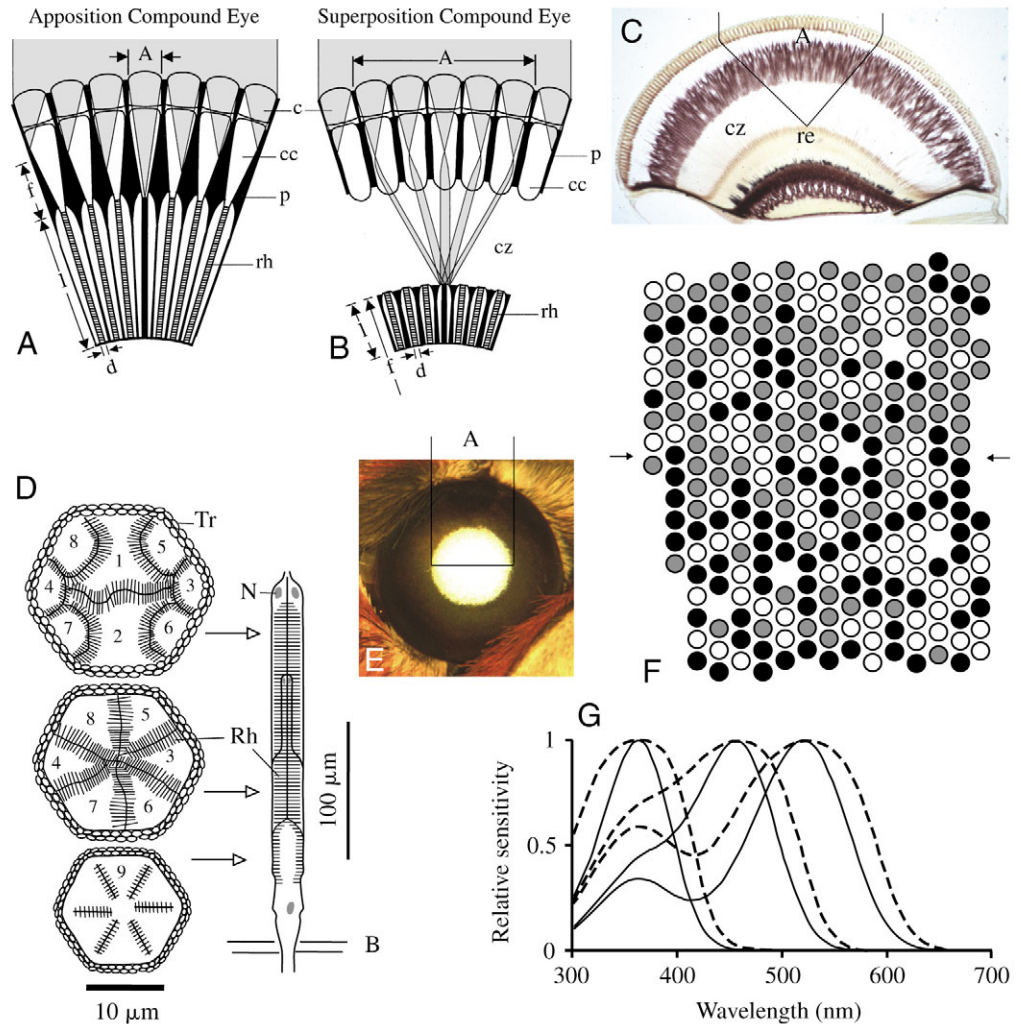
Animal groups known to lack the preconditions for colour vision include most deep-sea fish and crustaceans, most cephalopods, and some nocturnal and marine mammals (for references see Douglas et al., 1998; Kelber et al., 2003b). Even mammals with two types of photoreceptor (a rod and a cone) are most certainly colour-blind because they use one type, the rods, in dim light and the other type, the cones, in bright light. To prove that an animal uses colour vision, we need therefore to prove behaviourally that it is able to discriminate two colours by means of their spectral distributions and independent of their relative intensities (Kelber et al., 2003b).

Adaptations to nocturnal colour vision

A moonless night is about 100 million times darker than a day with bright sunshine (Fig. 1B). Still, a large number of animal species actively use their eyes at night, to find food, mates, or their homes. At nocturnal light levels, the quantal nature of light has severe consequences for vision. Quanta are distributed randomly and the noise, or uncertainty in an intensity measurement, is equivalent to \sqrt{n} quanta, when an average n quanta reach the eye at any one time interval. Discrimination of low contrasts is therefore severely impaired at low light intensities, and the eyes of nocturnal animals increase quantum capture by having large apertures and short focal lengths (for a review, see Warrant, 2004).

In lens eyes, high sensitivity is achieved by large pupils in combination with a short focal length. Examples of nocturnal animals with highly sensitive lens eyes include owl monkeys,

Fig. 2. (A) Schematic drawing of an apposition compound eye. The aperture (A) is defined by the diameter of a single corneal lens (c). Pigment (p) between the crystalline cones (cc) isolates ommatidia optically. Rhabdom (rh) length (l) and diameter (d) influence sensitivity; f, focal length. (B) Schematic drawing of a superposition compound eye. The clear zone (cz) interspaced between the crystalline cones and rhabdoms allows light entering through a large number of facets to be focussed on one rhabdom. This enlarges the aperture by factor of up to 1000. (C) Superposition eye of *Deilephila elpenor* (photo courtesy of Pär Brannström). cz, clear zone; re, retina. (D) Schematic drawings of the structure of the rhabdom of *D. elpenor* (adapted from Schlecht et al., 1978). B, basement membrane; Rh, rhabdom; Tr, tracheal tapetum, N, nucleus. Receptors 1 and 2 are blue- or UV-sensitive; receptors 3–9 are green-sensitive. (E) About 1000 facets of *D. elpenor* that build the superposition aperture glow when the eye is illuminated and viewed from the same direction. Photo courtesy of Michael Pfaff.



(F) Random arrays of ommatidial types in the crepuscular hawkmoth *Manduca sexta* (redrawn from White et al., 2003). White circles, ommatidia with all three receptor types; black circles, ommatidia with green and blue receptors; grey circles, ommatidia with green and UV receptors. Arrows show the eye horizon. (G) The effect of filtering in the long fused rhabdom of a dark-adapted nocturnal hawkmoth. The broken lines represent the sensitivities that would result from self-screening, if the long photoreceptors of *D. elpenor* had open rhabdoms. The solid lines represent the sensitivities resulting from filtering in the fused and tiered rhabdom.

owls, nocturnal geckos and many spiders. Large lenses with short focal lengths suffer from a severe optical problem: longitudinal chromatic aberration. Nocturnal vertebrates have probably solved this problem by having multifocal optics associated with slit pupils (Malmström and Kröger, 2006). Multifocal lenses focus light of different wavelengths in different spherical zones of the lens, thus producing a focussed image for multiple types of cone, and the slit pupil allows light to fall through all zones even when the pupil is closed (Kröger et al., 1999). However, even if this allows for colour vision in photopic and mesopic light intensities when cones are active, animals using a single type of rod in scotopic intensities remain colour-blind at night.

The compound eyes of insects and crustaceans come in two types (Land and Nilsson, 2002). In apposition compound eyes (Fig. 2A), each set of photoreceptors receives light only from its own tiny lens. These eyes are not very sensitive to light and

are not very well adapted to nocturnal vision, yet insects like grasshoppers and some nocturnal bees have driven these eyes to their extremes to cope with dim light levels: they possess large facet lenses, and wide and long rhabdoms (e.g. Warrant et al., 2004). Superposition compound eyes (Fig. 2B,C), possessed by nocturnal moths and beetles, are adapted to dim light vision. These eyes have a clear zone (marked cz in Fig. 2B,C) that allows light passing through a superposition aperture of up to 1000 lenses to be focussed onto one set of photoreceptors, thus enhancing the eye's sensitivity by a factor of up to 1000 (Fig. 2C,E). The photoreceptors of each rhabdom are enclosed by a tracheal tapetum (marked Tr in Fig. 2D) that contributes to the high sensitivity by doubling the light pass.

Besides optical adaptations, nocturnal animals have photoreceptors with large rhabdoms or outer segments (Figs 2D, 3A), and deep-sea fish have banded retinæ (for a review, see Warrant, 2004). In addition, many nocturnal eyes

have tapeta, mirrors behind the receptor layer that reflect light that is not absorbed during the first pass through the receptor and thus have the same effect as doubling the length of the receptor (Land and Nilsson, 2002). As a result of self-screening, long receptors have a much broader spectral sensitivity than short receptors (Warrant and Nilsson, 1998).

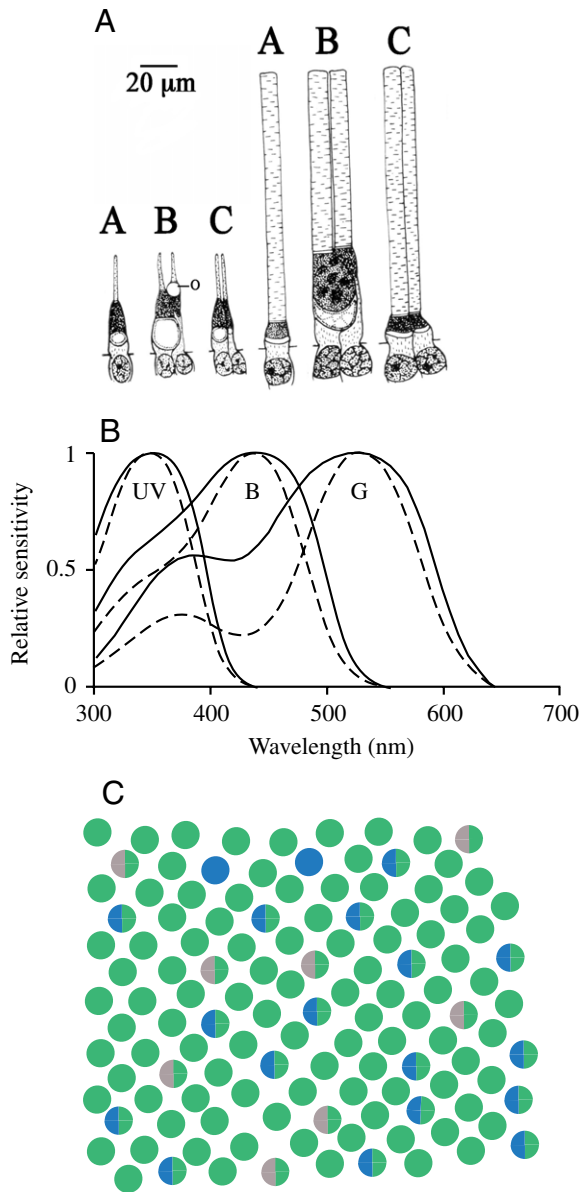


Fig. 3. (A) Anatomy of three anatomical types (A–C) of photoreceptor in diurnal (left) and nocturnal (right) geckos (modified from Underwood, 1970). Note the different lengths of the outer segments that cause different quantum captures. (B) Spectral sensitivities of the 5 μm short cones of a diurnal gecko (broken lines) and the 50 μm long cones of a nocturnal gecko (solid lines). UV, UV receptor; B, blue receptor; G, green receptor. (C) Cone mosaic of the nocturnal gecko *Teratoscincus scincus* (redrawn from Loew et al., 1996). Green circles, green-sensitive cones; blue circles, blue-sensitive cones; blue-green circles, double cones with a green and a blue receptor; grey-green circles, double cones with a green and a UV receptor.

They are more sensitive to light but the overlap between receptor sensitivities also makes them less useful for spectral discrimination and thus colour vision. Fig. 3B illustrates the difference between the sensitivities of dark-adapted gecko photoreceptors with outer segments of 5 μm and 50 μm length.

With very few photons available, pigment tuning also becomes essential, and should maximise signal-to-noise ratio (for recent discussions of this topic, see e.g. Douglas et al., 1998; Osorio and Vorobyev, 2005) for the visual system. Maximising photon catch and thus signal strength should solve half of the problem, but nocturnal moths, for example, have pigments sensitive to shorter wavelengths than those optimal for high photon catch (Johnsen et al., 2006). Visual pigments of nocturnal geckos are also sensitive to shorter wavelengths than those of related diurnal species (Ellingson et al., 1995; Loew et al., 1996), indicating that minimising noise is indeed an important factor.

Finally, temporal and spatial summation of receptor signals by neurons in the visual pathway can enhance the signal and improve signal-to-noise ratio considerably (Warrant, 1999; Warrant et al., 2004). In contrast, inhibitory interactions between receptors, including those involved in colour vision, lead to a lower signal-to-noise-ratio (Vorobyev, 1997). A single long-wavelength receptor has, under most nocturnal conditions, a higher sensitivity than either a short-wavelength-sensitive receptor or any combination of both (Kelber et al., 2003a; Osorio and Vorobyev, 2005; Johnsen et al., 2006). With very few photons available, a monochromatic eye with an optimally tuned pigment can thus discriminate more shades of colour simply by means of intensity contrast than can a dichromatic or trichromatic eye (Vorobyev, 1997). Many animals, including ourselves, therefore sacrifice colour vision at night (Dusenbery, 1992). However, as demonstrated below, spectral information is just as useful at night as it is during day (Kelber et al., 2002; Land and Osorio, 2003; Johnsen et al., 2006), and some animals capture enough photons to see colours even in dim light.

Nocturnal colour vision for object constancy

How useful colour information can be, compared to the achromatic signal provided by a single type of receptor, is easily demonstrated. One of the most general tasks that animals use vision for is recognition of food items, hosts, mates, landmarks, nests and other objects. As the visual appearance of objects, changes dramatically with changing illumination, however, mechanisms for object constancy are extremely important. Fig. 4 shows how a yellow flower, a blue flower and a typical leaf background change contrast and colour between two illuminations measured at the same place, within less than a minute: direct sunlight and a shadow cast by an observer. The contrasts of both colours to the green background (Fig. 4C) change dramatically; the blue flower looks brighter than the yellow flower in the shadow but darker in direct sunlight. However, in the colour space of an insect (Fig. 4D), both

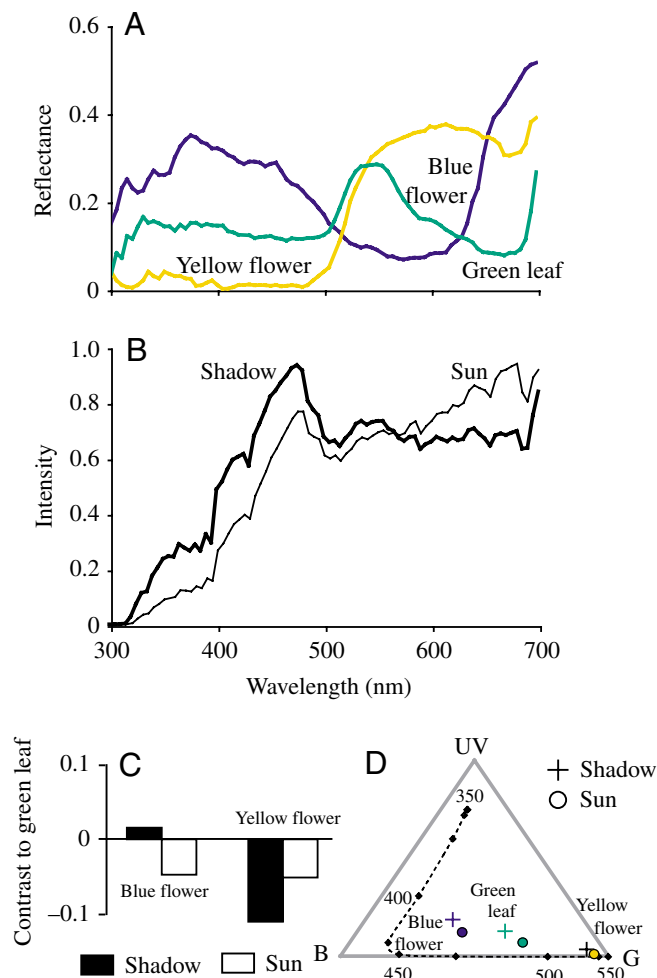


Fig. 4. The use of chromatic vision. (A) The reflectances of a typical yellow and a typical blue flower and a green leaf. (B) The spectral composition of direct sunlight and skylight in a shadow, measured shortly before sunset, on a summer day in Utah, USA. (C) The achromatic contrast between both flowers and the green leaf in both illuminations differ dramatically for the green receptor of *D. elpenor*. (D) The colour loci of all three stimuli in the colour triangle of *D. elpenor* are rather constant in both illuminations, even without the assumption of colour constancy. UV, UV light; B, blue light; G, green light. (For details and methods of all calculations, see Johnsen et al., 2006.)

colours can easily be separated under both illuminations, even without any assumption of colour constancy. The colour triangle shown here is a two-dimensional projection of the three-dimensional colour space of an insect with three receptor types. It disregards intensity. It shows that colour is a much more reliable and constant property of objects than achromatic contrast. Obviously, the same applies in dim light. The colour of light changes dramatically during dawn and dusk, and between moonlight and starlight, and colour vision allows animals to identify objects more reliably under these conditions (Johnsen et al., 2006). The colour signal changes much less with changing illumination than the achromatic signal (Johnsen et al., 2006).

Scotopic colour vision in crepuscular and nocturnal hawkmoths

Most hawkmoths are active during twilight (crepuscular) or at night (nocturnal). They have superposition compound eyes with a large superposition aperture and tracheal tapeta (Fig. 2B–D), and are therefore well adapted to nocturnal vision. Similar to the well-known honeybee, they rely on one single set of three spectral receptor types that are maximally sensitive to ultraviolet, blue and green light, for vision at all light intensities (Schwemer and Paulsen, 1973). As a result of self-screening, their long photoreceptors should have broadened sensitivity functions when they are dark-adapted (broken curve in Fig. 2G; Warrant and Nilsson, 1998; Kelber et al., 2003a). However, the rhabdomeres of moth photoreceptors are fused and work as spectral filters for each other, thus narrowing the sensitivity functions and making them well suited for colour vision (solid curve in Fig. 2G; for formulae to calculate the curves see Johnsen et al., 2006). Diurnal flower visitors such as honeybees have long been known to use colour to detect, recognise and discriminate the colours of rewarding flowers (Frisch, 1914).

Using a method developed by von Frisch in 1914, we have recently demonstrated colour vision in the crepuscular and nocturnal hawkmoths *Deilephila elpenor*, *Hyles lineata* and *Hyles gallii* (von Frisch, 1914; Kelber et al., 2002; Kelber et al., 2003a). The moths were trained to associate a reward of sucrose solution with the blue or yellow colour of artificial flowers (Fig. 5A). After training, they were able to discriminate the training colour from eight different shades of grey and from two other colours (Fig. 5B,C). As the different shades of grey provided different achromatic cues to the moths, they could only rely on the chromatic signal (or colour) for the discrimination. They were therefore unable to discriminate the training blue (or yellow) from a lighter or darker shade of blue (or yellow, respectively), because these stimuli differed only in the achromatic and not the chromatic signal (right panels in Fig. 5B,C). The results shown in Fig. 5 were obtained at a light intensity similar to light levels on a starlit night (0.0001 cd m^{-2}). Moths also discriminated between two colours looking white to the human eye, one absorbing ultraviolet light and one reflecting it (Kelber et al., 2002). This proves that the nocturnal colour vision of *D. elpenor* also extends into the ultraviolet range. In addition, *D. elpenor* has good colour constancy and is able to recognize rewarding flowers under changed illumination (Balkenius and Kelber, 2004).

How can moths achieve colour vision at light intensities where humans are colour-blind? The number of photons captured by the receptors in each ommatidium, at starlight intensities, ranges between 1 and 25 photons, per receptor channel and receptor integration time. With these numbers of photons, the noise level (\sqrt{N} equivalent to between 1 and 5 photons) would make discrimination impossible. We therefore have to propose that nocturnal moths use spatial and/or temporal summation (Warrant, 1999) to improve the signal-to-noise ratio and allow the colour discrimination we observed.

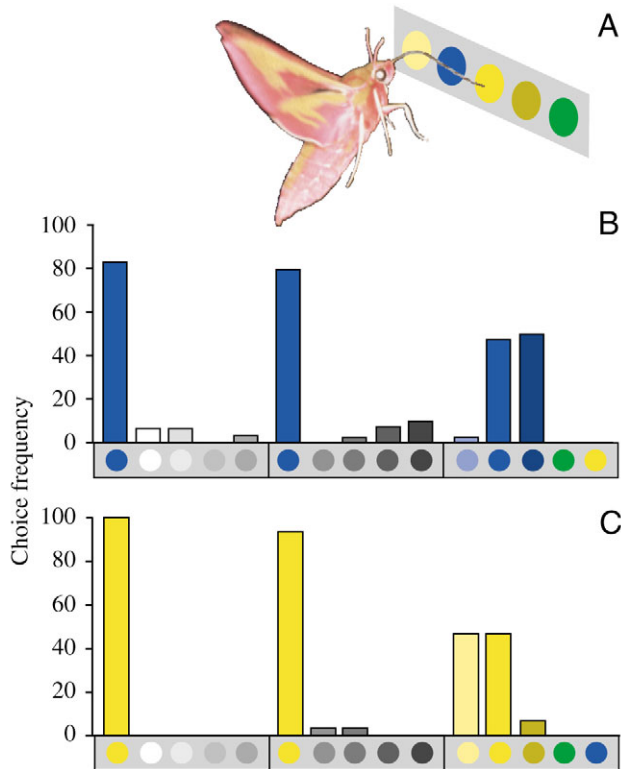


Fig. 5. At starlight intensities of illumination, nocturnal hawkmoths of the species *Deilephila elpenor* learned to discriminate a training colour from eight different shades of grey and from two other colours but not from brighter or darker shades of the training colour. (A) The animal choosing one of the stimuli in the set-up. Discrimination occurred when the training colour was blue (B) or yellow (C). (Data from Kelber et al., 2002.)

Calculations show that by having three spectral types of receptor, the hawkmoths sacrifice absolute sensitivity, and this indicates that colour vision is highly relevant for them, just as it is for diurnal flower visitors (Kelber et al., 2003a; Johnsen et al., 2006).

Colour vision in nocturnal geckos

Among vertebrates, lizards have lost their dual retina during evolution, and are now left only with cones (Underwood, 1970). Nocturnal geckos have evolved from diurnal lizards and thus have a similar retina: no rods but three spectral types of cone sensitive to ultraviolet, blue and green light (360, 440 and 520 nm, respectively, Fig. 3B,C) (Loew et al., 1996). Adaptations to the nocturnal life-style include very long cone outer segments (Fig. 3A) and a combination of large pupil and short focal length. Since this condition results in severe chromatic aberration it is assumed that geckos may have multifocal lens optics (Kröger et al., 1999). Being ambush predators, geckos use motion vision to detect prey, and motion vision tends to be colour-blind in most animals (Kelber et al., 2003b). Are geckos nonetheless able to use colour as a cue?

It is difficult to convince geckos to use colour cues but it

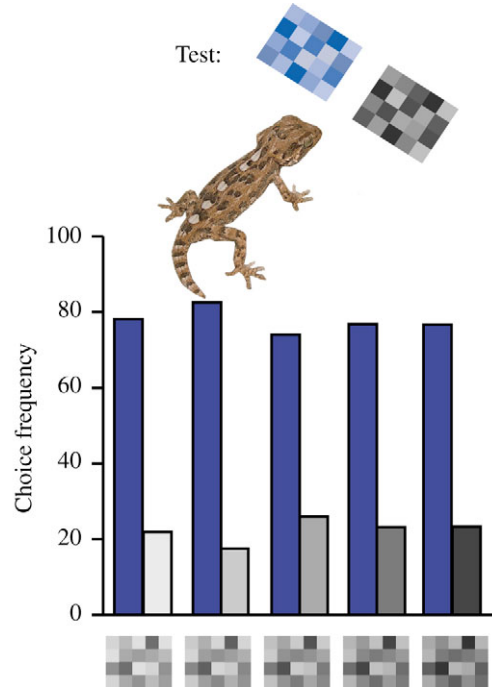


Fig. 6. Two nocturnal geckos of the species *Tarentola chezialiae* learnt to take a cricket from forceps decorated with a pattern made from blue squares (right). The forceps decorated with a grey pattern (left) always held a salted cricket, which geckos always refused. Both geckos predominantly chose blue in all tests, independent of the overall intensity of the grey pattern (represented by the different shades of the grey patterns on the abscissa). (Data from Roth and Kelber, 2004.)

was possible using a method similar to that used with nocturnal hawkmoths (Roth and Kelber, 2004). A well-tasting cricket was presented in front of a blue pattern, and a salty (badly-tasting) cricket in front of a grey pattern (inset in Fig. 6). The shades of grey and blue were made equally bright for the geckos, thus making discrimination by means of achromatic cues very unlikely. To make intensity completely unreliable for the geckos, we varied it between training trials leaving only colour as the signal to be learned. Two geckos learned to discriminate between both colours, at light intensities mimicking dim moonlight (Fig. 6; 0.002 cd m^{-2}). They might well be able to use colour even at lower light intensities. Their anatomical adaptations thus allow nocturnal geckos to use cone-based colour vision at night (Roth and Kelber, 2004). The question as to why a predator should preserve and use colour vision, even in dim light, remains to be answered. Most probably, object constancy is important even for predators.

Bioluminescence and colour vision

Glow worms and fireflies, and many fishes, crustaceans and cephalopods living in the deep sea, produce their own light to be seen in the darkness: bioluminescent signals. They are bright light sources, the intensity of which should make detection a simple task. It is therefore commonly thought that

bioluminescent communication is based on achromatic matched filtering: the spectral sensitivity of the eye is matched to the mate's signal, and the temporal patterns of flashes are species-specific. However, in fireflies (*Photinus* sp.) and glow-worms (*Lampyrus noctiluca*), there is good evidence that colour vision is involved. In fireflies, behavioural data (Lall and Worthy, 2000) can only be explained by assuming a chromatic interaction between the blue and green receptors. Recent experiments prove this to be the case for the European glow-worm *Lampyrus noctiluca* (Booth et al., 2004). The Central-American click beetle *Pyrophorus plagiophthalmus* (Elateridae) has two dorsal light organs emitting green light and a third ventral organ emitting orange light (Stolz et al., 2003). Behavioural data are missing, but we may speculate on their use of colour.

Behavioural evidence is also lacking for firefly squids, but it is known that these molluscs and some deep-sea crustaceans and fish possess two receptor types that may allow them to discriminate between the colour of the down-dwelling daylight and the colour of their bioluminescence (for references see Kelber, in press; Douglas et al., 1998).

Some deep-sea dragon fish have two-coloured bioluminescence in the blue and far-red, and photoreceptors sensitive to the red fluorescence (Douglas et al., 1998). Whether they compare signals from this red receptor and the blue-sensitive rod for colour vision or just use the red receptor as a parallel channel may remain their secret since these animals are difficult to access and hard if not impossible to study in behavioural tests (Douglas et al., 1998). In general, however, bioluminescent stimuli are bright, and the eyes of these animals do not have to be quite as sensitive as those of nocturnal moths or geckos.

Other species might join the club

Are moths and geckos special or do more species have the capability to see colour at night? We can assume that other large nocturnal insects with superposition eyes, including large moths and beetles, should be able to see colour in dim light. Even insects with highly sensitive apposition eyes, including grasshoppers and large nocturnal bees such as the carpenter bee *Xylocopa (Nyctimelitta) proximata* that forages on moonless nights (Somanathan and Borges, 2001; H. Somanathan and R. Borges, unpublished observations), and the Halictid bee *Megalopta genalis* that forages during dawn and dusk under the canopy of the tropical rain forest (Warrant et al., 2004), might possess this ability, but this remains to be studied. Large arthropods with lens eyes and three types of photoreceptors in the retina, such as the nocturnal wandering spider *Cupiennius salei* (Walla et al., 1996), might also have colour vision in dim light.

Among the vertebrates, another nocturnal group besides the geckos might see colour at night. In contrast to other vertebrates, toads and frogs have two types of rod (Liebman and Entine, 1968), maximally sensitive to 432 nm and 502 nm, respectively. They use colour vision for mate recognition in bright light (for references, see Kelber et al., 2003) and might

use rod-based colour vision for the same purpose in dim light. Alternatively, they might only use the achromatic signal that results from summing the signals in both rods types. The latter has been shown to be the case for the optomotor response in the green treefrog *Hyla cinerea* (King et al., 1993). Finally, in some fish, the red-sensitive cones are more sensitive to light than the other cone types and can interact with rods (Roessel et al., 1997). This has been proved by classical conditioning experiments in anaesthetised goldfish (Powers and Easter, 1978) and it might apply to other species as well. It is, however, unknown whether and how the chromatic signal derived from rods and red cones is used by the fish.

Conclusions

Colour vision is just as useful at night as it is during the day. The reason for this probably lies in the dramatic changes in the colour of light that animals experience both between sun- or moonlit areas and shadows, and during the twilight period. Animals with highly sensitive eyes can see colour in very dim light, if their eyes have the preconditions: several types of receptor that are active simultaneously. Direct evidence for nocturnal colour vision only exists for nocturnal hawkmoths and geckos, but more species need to be studied. Rather than asking why some animals do have colour vision at night, we may have to ask why many others, among them most vertebrates, do not!

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